

APPLICATION UNDER UNITED STATES PATENT LAWS

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Invention: Chemical Compounds

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This is a:

- ☐ Provisional Application
- ☒ Regular Utility Application
- ☐ Continuing Application
☐ The contents of the parent are incorporated by reference
- ☐ PCT National Phase Application
- ☐ Design Application
- ☐ Reissue Application
- ☐ Plant Application
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Sub. Spec. Filed _____
in App. No. _____ / _____
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SPECIFICATION

SUBSTANTIVE SPECIFICATION

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Specification for use in: US

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CHEMICAL COMPOUNDS

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CHEMICAL COMPOUNDS

This invention relates to polymorphisms in the human OATPC gene and corresponding novel allelic polypeptides encoded thereby. The invention also relates to methods and materials for analysing allelic variation in the OATPC gene, and to the use of OATPC polymorphism in treatment of diseases with OATPC transportable drugs.

Na⁺-independent organic anion transporting polypeptide (OATP) C gene is a member of the OATP supergene family involved in multifunctional transport of organic anion. OATPC transports the organic anion taurocholate, conjugated steroids: DHEAS, estradiol 17 β -D-glucuronide and estrone-3-sulfate, eicosanoids: PGE₂, thromboxane B₂, leukotriene C₄, and E₄, and thyroid hormones T₄ and T₃^{1,2}. OATPC has also been shown to be involved in the transport of xenobiotics, and drugs involved in lipid lowering *e.g.* statins¹. Statins have been referred to as a first-line therapy for patients with atherosclerotic vascular diseases. The OATPC gene and its product is also thought to be of importance in other diseases due to its transport of DHEAS an adrenal steroid which has been suggested to have positive neuropsychiatric, immune, and metabolic effects³. Due to the substrate specificity, location in the liver, and being exclusively expressed in the liver, Abe *et al* suggested that OATPC could be the predominant clearance mechanism of several endogenous and exogenous substrates in the liver. OATPC is the first human molecule reported to transport thyroid hormones².

This liver specific transporter may be useful in liver-specific drug delivery systems and liver-specific chemotherapy, bile acid formation and the pathogenesis of diseases such as cholestasis, hyperbilirubinemia and thyroid hormone resistance.

The OATPC gene (sometimes called OAPT2 in the literature) has been cloned by four different groups, annotated and published as EMBL accession numbers AB026257 (OATPC, 2452bp), AF205071(OATP2, 2830, ref 1), AJ132573(OATP2, 2778)⁴, and AF060500 (LST-

¹ A Novel Human Hepatic Organic Anion Transporting Polypeptide (OATP2), Hsiang *et al* J Biol Chem **274**, 37161-37168 (1999)

² Identification of a Novel Gene Family Encoding Human Liver-specific Organic Anion Transporter LST-1, Takaaki Abe *et al* J Biol Chem **274**, 17159-17163 (1999)

³ Bates *et al* (1998) Curr. Opin. Endocrinol. Diab. **5**, 357-366

⁴ A novel human organic anion transporting polypeptide localised to the basolateral hepatocyte membrane, König Jorg *et al* (2000) Am J Physiol. Gastrointest. Liver Physiol. **278**: G156-G164

1)². Polymorphism has been reported by Tamai⁵ which is Asn130Asp and Val174Ala although any functional effect was stated therein to be not clear. Konig (2000) J Biol Chem 275, 23161-23168 describes the genomic organisation of OATP 1, 2 and 8. International patent application WO 00/08157 describes human anion transporter genes and some

5 polymorphisms.

All positions herein of polymorphisms in the OATPC polynucleotide relate to the position in one of SEQ ID NO 1 or 3-12 unless stated otherwise or apparent from the context.

All positions herein of polymorphisms in the OATPC polypeptide relate to the position in SEQ ID NO 2 unless stated otherwise or apparent from the context.

10 One approach is to use knowledge of polymorphisms to help identify patients most suited to therapy with particular pharmaceutical agents (this is often termed "pharmacogenetics"). Pharmacogenetics can also be used in pharmaceutical research to assist the drug selection process. Polymorphisms are used in mapping the human genome and to elucidate the genetic component of diseases. The reader is directed to the following
15 references for background details on pharmacogenetics and other uses of polymorphism detection: Linder *et al.* (1997), Clinical Chemistry, **43**, 254; Marshall (1997), Nature Biotechnology, **15**, 1249; International Patent Application WO 97/40462, Spectra Biomedical; and Schafer *et al.* (1998), Nature Biotechnology, **16**, 33.

20 Clinical trials have shown that patient response to treatment with pharmaceuticals is often heterogeneous. Thus there is a need for improved approaches to pharmaceutical agent design and therapy.

Point mutations in polypeptides will be referred to as follows: natural amino acid (using 1 or 3 letter nomenclature), position, new amino acid. For (a hypothetical) example
25 "D25K" or "Asp25Lys" means that at position 25 an aspartic acid (D) has been changed to lysine (K). Multiple mutations in one polypeptide will be shown between square brackets with individual mutations separated by commas.

The present invention is based on the discovery of polymorphisms in OATPC.

According to one aspect of the present invention there is provided a method for the detection of a polymorphism in OATPC in a human, which method comprises determining the
30 sequence of the human at at least one polymorphic position and determining the status of the

⁵ Tamai *et al* (2000), BBRC, **273**, 251-60

human by reference to polymorphism in the OATPC gene. Preferred polymorphic positions are one or more of the following positions:

A method for the detection of a polymorphism in OATPC in a human, which method comprises determining the sequence of the human at at least one of the following polymorphic

5 positions:

positions 510, 696, 1299, 1312, 1347, 1561, 2028, 2327 and 2342 in sequence of the OATPC gene as defined by the position in SEQ ID NO: 1;

positions 400, 405, 488 and 643 in OATPC polypeptide defined by position in SEQ ID NO: 2;

positions 321 and 1332 defined by position in SEQ ID NO 3;

10 position 41 defined by position in SEQ ID NO 4;

positions 109 and 244 defined by position in SEQ ID NO 5;

positions 117 and 283 defined by position in SEQ ID NO 6;

positions 209 and 211 defined by position in SEQ ID NO 7;

positions 63 to 68 defined by position in SEQ ID NO 8;

15 position 53 defined by position in SEQ ID NO 9;

position 75 defined by position in SEQ ID NO 10;

position 162 defined by position in SEQ ID NO 11; and

positions 84 defined by position in SEQ ID NO 12.

The term human includes both a human having or suspected of having a OATPC
20 mediated disease and an asymptomatic human who may be tested for predisposition or susceptibility to such disease. At each position the human may be homozygous for an allele or the human may be a heterozygote.

The term polymorphism includes single nucleotide substitution, nucleotide insertion and nucleotide deletion which in the case of insertion and deletion includes insertion or
25 deletion of one or more nucleotides at a position of a gene.

The method for diagnosis is preferably one in which the sequence is determined by a method selected from amplification refractory mutation system and restriction fragment length polymorphism.

The status of the individual may be determined by reference to allelic variation at any
30 one, two, three, four, five, six, seven, eight, nine or more positions.

The test sample of nucleic acid is conveniently a sample of blood, bronchoalveolar lavage fluid, sputum, or other body fluid or tissue obtained from an individual. It will be

appreciated that the test sample may equally be a nucleic acid sequence corresponding to the sequence in the test sample, that is to say that all or a part of the region in the sample nucleic acid may firstly be amplified using any convenient technique e.g. PCR, before analysis of allelic variation.

- 5 It will be apparent to the person skilled in the art that there are a large number of analytical procedures which may be used to detect the presence or absence of variant nucleotides at one or more polymorphic positions of the invention. In general, the detection of allelic variation requires a mutation discrimination technique, optionally an amplification reaction and optionally a signal generation system. Table 1 lists a number of mutation
- 10 detection techniques, some based on the PCR. These may be used in combination with a number of signal generation systems, a selection of which is listed in Table 2. Further amplification techniques are listed in Table 3. Many current methods for the detection of allelic variation are reviewed by Nollau *et al.*, Clin. Chem. **43**, 1114-1120, 1997; and in standard textbooks, for example "Laboratory Protocols for Mutation Detection", Ed. by U.
- 15 Landegren, Oxford University Press, 1996 and "PCR", 2nd Edition by Newton & Graham, BIOS Scientific Publishers Limited, 1997.

Abbreviations:

ALEX™	Amplification refractory mutation system linear extension
APEX	Arrayed primer extension
ARMST™	Amplification refractory mutation system
b-DNA	Branched DNA
bp	base pair
CMC	Chemical mismatch cleavage
COPS	Competitive oligonucleotide priming system
DGGE	Denaturing gradient gel electrophoresis
FRET	Fluorescence resonance energy transfer
HMG-CoA	3-hydroxy-3-methylglutaryl-coenzyme A
LCR	Ligase chain reaction
MASDA	Multiple allele specific diagnostic assay
NASBA	Nucleic acid sequence based amplification
OATP	Na ⁺ -independent organic anion transporting polypeptide
OLA	Oligonucleotide ligation assay
PCR	Polymerase chain reaction
PTT	Protein truncation test
RFLP	Restriction fragment length polymorphism
SDA	Strand displacement amplification
SNP	Single nucleotide polymorphism
SSCP	Single-strand conformation polymorphism analysis

SSR	Self sustained replication
TGGE	Temperature gradient gel electrophoresis

Table 1 - Mutation Detection Techniques

General: DNA sequencing, Sequencing by hybridisation

Scanning: PTT*, SSCP, DGGE, TGGE, Cleavase, Heteroduplex analysis, CMC, Enzymatic

5 mismatch cleavage

* Note: not useful for detection of promoter polymorphisms.

Hybridisation Based

Solid phase hybridisation: Dot blots, MASDA, Reverse dot blots,

Oligonucleotide arrays (DNA Chips)

10 Solution phase hybridisation: Taqman™ - US-5210015 & US-5487972 (Hoffmann-La Roche), Molecular Beacons - Tyagi *et al* (1996), Nature Biotechnology, 14, 303; WO 95/13399 (Public Health Inst., New York)

Extension Based: ARMST™, ALEX™ - European Patent No. EP 332435 B1 (Zeneca Limited), COPS - Gibbs *et al* (1989), Nucleic Acids Research, 17, 2347.

15 **Incorporation Based:** Mini-sequencing, APEX

Restriction Enzyme Based: RFLP, Restriction site generating PCR

Ligation Based: OLA

Other: Invader assay

20 Table 2 - Signal Generation or Detection Systems

Fluorescence: FRET, Fluorescence quenching, Fluorescence polarisation - United Kingdom Patent No. 2228998 (Zeneca Limited)

Other: Chemiluminescence, Electrochemiluminescence, Raman, Radioactivity, Colorimetric, Hybridisation protection assay, Mass spectrometry

25

Table 3 - Further Amplification Methods

SSR, NASBA, LCR, SDA, b-DNA

Preferred mutation detection techniques include ARMST™, ALEX™, COPS, Taqman,

30 Molecular Beacons, RFLP, and restriction site based PCR and FRET techniques.

Particularly preferred methods include ARMSTTM and RFLP based methods. ARMSTTM is an especially preferred method.

In a further aspect, the diagnostic methods of the invention are used to assess the pharmacogenetics of a drug transportable by OATPC.

5 Assays, for example reporter-based assays, may be devised to detect whether one or more of the above polymorphisms affect transcription levels and/or message stability.

Individuals who carry particular allelic variants of the OATPC gene may therefore exhibit differences in their ability to regulate protein biosynthesis under different physiological conditions and will display altered abilities to react to different diseases. In
10 addition, differences arising as a result of allelic variation may have a direct effect on the response of an individual to drug therapy. The diagnostic methods of the invention may be useful both to predict the clinical response to such agents and to determine therapeutic dose.

In a further aspect, the diagnostic methods of the invention, are used to assess the predisposition and/or susceptibility of an individual to diseases mediated by OATPC. This
15 may be particularly relevant in the development of hyperlipoproteinemia and cardiovascular disease and the present invention may be used to recognise individuals who are particularly at risk from developing these conditions.

In a further aspect, the diagnostic methods of the invention are used in the development of new drug therapies which selectively target one or more allelic variants of the
20 OATPC gene. Identification of a link between a particular allelic variant and predisposition to disease development or response to drug therapy may have a significant impact on the design of new drugs. Drugs may be designed to regulate the biological activity of variants implicated in the disease process whilst minimising effects on other variants.

In a further diagnostic aspect of the invention the presence or absence of variant
25 nucleotides is detected by reference to the loss or gain of, optionally engineered, sites recognised by restriction enzymes.

According to another aspect of the present invention there is provided a human OATPC gene or its complementary strand comprising a variant allelic polymorphism at one or more of positions defined herein or a fragment thereof of at least 20 bases comprising at least
30 one novel polymorphism.

Fragments are at least 17 bases, more preferably at least 20 bases, more preferably at least 30 bases.

According to another aspect of the present invention there is provided a polynucleotide comprising at least 20 bases of the human OATPC gene and comprising an allelic variant selected from any one of the following:

Region	variant	Position in SEQ ID NO	SEQ ID NO
Exon 4	A	510	1
Exon 5	T	670	1
Exon 5	T	696	1
Exon 9	G	1299	1
Exon 9	A	1312	1
Exon 9	A	1347	1
Exon 10	C	1561	1
Exon 14	C	2028	1
3'UTR	Insert T	2327	1
3'UTR	C	2342	1
Promoter	G	321	3
Promoter	C	1332	3
Intron 1	A	41	4
Intron 2	G	109	5
Intron 2	G	244	5
Intron 3	A	117	6
Intron 3	A	283	6
Intron 4	A	209	7
Intron 4	A	211	7
Intron 4	Deletion CTTGTA	63	8
Intron 6	T	53	9
Intron 9	Insert TTC	75	10
Intron 11	Insert T	162	11
Intron 12	C	84	12

- 5 According to another aspect of the present invention there is provided a human OATPC gene or its complementary strand comprising a polymorphism, preferably corresponding with one or more the positions defined herein or a fragment thereof of at least 20 bases comprising at least one polymorphism.

Fragments are at least 17 bases, more preferably at least 20 bases, more preferably at 10 least 30 bases.

The invention further provides a nucleotide primer which can detect a polymorphism of the invention.

According to another aspect of the present invention there is provided an allele specific primer capable of detecting a OATPC gene polymorphism, preferably at one or more of the positions as defined herein.

5 An allele specific primer is used, generally together with a constant primer, in an amplification reaction such as a PCR reaction, which provides the discrimination between alleles through selective amplification of one allele at a particular sequence position e.g. as used for ARMSTTM assays. The allele specific primer is preferably 17- 50 nucleotides, more preferably about 17-35 nucleotides, more preferably about 17-30 nucleotides.

10 An allele specific primer preferably corresponds exactly with the allele to be detected but derivatives thereof are also contemplated wherein about 6-8 of the nucleotides at the 3' terminus correspond with the allele to be detected and wherein up to 10, such as up to 8, 6, 4, 2, or 1 of the remaining nucleotides may be varied without significantly affecting the properties of the primer.

15 Primers may be manufactured using any convenient method of synthesis. Examples of such methods may be found in standard textbooks, for example "Protocols for Oligonucleotides and Analogues; Synthesis and Properties," Methods in Molecular Biology Series; Volume 20; Ed. Sudhir Agrawal, Humana ISBN: 0-89603-247-7; 1993; 1st Edition. If required the primer(s) may be labelled to facilitate detection.

20 According to another aspect of the present invention there is provided an allele-specific oligonucleotide probe capable of detecting a OATPC gene polymorphism, preferably at one or more of the positions defined herein.

The allele-specific oligonucleotide probe is preferably 17- 50 nucleotides, more preferably about 17-35 nucleotides, more preferably about 17-30 nucleotides.

25 The design of such probes will be apparent to the molecular biologist of ordinary skill. Such probes are of any convenient length such as up to 50 bases, up to 40 bases, more conveniently up to 30 bases in length, such as for example 8-25 or 8-15 bases in length. In general such probes will comprise base sequences entirely complementary to the corresponding wild type or variant locus in the gene. However, if required one or more mismatches may be introduced, provided that the discriminatory power of the oligonucleotide 30 probe is not unduly affected. The probes of the invention may carry one or more labels to facilitate detection.

According to another aspect of the present invention there is provided an allele specific primer or an allele specific oligonucleotide probe capable of detecting a OATPC gene polymorphism at one of the positions defined herein.

According to another aspect of the present invention there is provided a diagnostic kit
5 comprising an allele specific oligonucleotide probe of the invention and/or an allele-specific primer of the invention.

The diagnostic kits may comprise appropriate packaging and instructions for use in the methods of the invention. Such kits may further comprise appropriate buffer(s) and polymerase(s) such as thermostable polymerases, for example taq polymerase.

10 In another aspect of the invention, the single nucleotide polymorphisms of this invention may be used as genetic markers in linkage studies. This particularly applies to the polymorphisms of relatively high frequency. The OATPC gene is on chromosome 12p (as shown from a database search with the cDNA as a query sequence). Low frequency polymorphisms may be particularly useful for haplotyping as described below. A haplotype is
15 a set of alleles found at linked polymorphic sites (such as within a gene) on a single (paternal or maternal) chromosome. If recombination within the gene is random, there may be as many as 2^n haplotypes, where 2 is the number of alleles at each SNP and n is the number of SNPs. One approach to identifying mutations or polymorphisms which are correlated with clinical response is to carry out an association study using all the haplotypes that can be identified in
20 the population of interest. The frequency of each haplotype is limited by the frequency of its rarest allele, so that SNPs with low frequency alleles are particularly useful as markers of low frequency haplotypes. As particular mutations or polymorphisms associated with certain clinical features, such as adverse or abnormal events, are likely to be of low frequency within the population, low frequency SNPs may be particularly useful in identifying these mutations
25 (for examples see: Linkage disequilibrium at the cystathionine beta synthase (CBS) locus and the association between genetic variation at the CBS locus and plasma levels of homocysteine. *Ann Hum Genet* (1998) **62**:481-90, De Stefano V, Dekou V, Nicaud V, Chasse JF, London J, Stansbie D, Humphries SE, and Gudnason V; and Variation at the von willebrand factor (vWF) gene locus is associated with plasma vWF:Ag levels: identification of three novel
30 single nucleotide polymorphisms in the vWF gene promoter. *Blood* (1999) **93**:4277-83, Keightley AM, Lam YM, Brady JN, Cameron CL, Lillicrap D).

According to another aspect of the present invention there is provided a computer readable medium comprising at least one novel sequence of the invention stored on the medium. The computer readable medium may be used, for example, in homology searching, mapping, haplotyping, genotyping or pharmacogenetic analysis.

5 According to another aspect of the present invention there is provided a method of treating a human in need of treatment with a drug transportable by OATPC in which the method comprises:

i) detection of a polymorphism in OATPC in the human, which detection comprises determining the sequence of the human at one or more of the following positions:

10 positions 487, 510, , 554, 670, 696, 819, 820, 1299, 1312, 1347, 1561, 2028, 2327 and 2342

in sequence of the OATPC gene as defined by the position in SEQ ID NO: 1;

positions 130, 152, 174, 241, 400, 405, 488 and 643 in OATPC polypeptide defined by position in SEQ ID NO: 2;

positions 321 and 1332 defined by position in SEQ ID NO 3;

15 position 41 defined by position in SEQ ID NO 4;

positions 109 and 244 defined by position in SEQ ID NO 5;

positions 117 and 283 defined by position in SEQ ID NO 6;

positions 209 and 211 defined by position in SEQ ID NO 7;

positions 63 to 68 defined by position in SEQ ID NO 8;

20 position 53 defined by position in SEQ ID NO 9;

position 75 defined by position in SEQ ID NO 10;

position 162 defined by position in SEQ ID NO 11; and

positions 84 defined by position in SEQ ID NO 12.

and determining the status of the human by reference to polymorphism in the OATPC gene;

25 and

ii) administering an effective amount of the drug.

Preferably determination of the status of the human is clinically useful. Examples of clinical usefulness include deciding which statin drug or drugs to administer and/or in deciding on the effective amount of the statin drug or drugs. Statins already approved for use
30 in humans include atorvastatin, cerivastatin, fluvastatin, pravastatin and simvastatin. The reader is referred to the following references for further information: Drugs and Therapy Perspectives (12th May 1997), 9: 1-6; Chong (1997) Pharmacotherapy 17: 1157-1177; Kellick

(1997) Formulary **32**: 352; Kathawala (1991) Medicinal Research Reviews, **11**: 121-146; Jahng (1995) Drugs of the Future **20**: 387-404, and Current Opinion in Lipidology, (1997), **8**, 362 - 368. A preferred statin drug is compound 3a (S-4522) in Watanabe (1997) Bioorganic and Medicinal Chemistry **5**: 437-444; now called rosuvastatin, see Olsson (2001) American Journal of Cardiology, **87**, supplement 1, 33-36. The term "drug transportable by OATPC" means that transport by OATPC in humans is an important part of a drug exerting its pharmaceutical effect in man. For example, some statins have to be transported to the liver by OATPC to exert their lipid lowering effects.

According to another aspect of the present invention there is provided a method of treating a human in need of treatment with a drug transportable by OATPC in which the method comprises:

- i) diagnosis of a single nucleotide polymorphism in OATPC gene in the human, which diagnosis preferably comprises determining the sequence of the nucleic acid at position 487 in the coding sequence of the OATPC gene as defined by the position in SEQ ID NO: 1.
- and determining the status of the human by reference to polymorphism in the OATPC gene;
- and
- ii) administering an effective amount of the drug.

Although the polymorphism at position 487 (A→G; Asn130Asp) was reported by Tamai, no functional effect was attributed. Indeed the following polymorphisms are known but have not previously been attributed a role in statin pharmacogenetics as disclosed herein.

OATPC polymorphisms

Position of nucleotide as in sequence 1 of OATPC patent

Position of amino acid as in sequence 2 of OATPC patent

Exon	nucleotide	SNP	Aminoacid	Change	Change
4	487	A>G	130	Asn>Asp	N>D
4	554	G>A	152	Arg>Lys	R>K
5	620	T>C	174	Val>Ala	V>A
5	670	C>T	191	Leu	L
6	819	A>T	240	Val	V
6	820	G>A	241	Asp>Asn	D>N

According to another aspect of the present invention there is provided use of a drug transportable by OATPC in preparation of a medicament for treating a disease in a human

diagnosed as having a polymorphism therein, preferably at one or more of the positions defined herein. Preferably the disease is cardiovascular.

According to another aspect of the present invention there is provided a pharmaceutical pack comprising OATPC transportable drug and instructions for administration of the drug to humans diagnostically tested for a single nucleotide polymorphism therein, preferably at one or more of the positions defined herein.

According to another aspect of the present invention there is provided an allelic variant of human OATPC polypeptide comprising at least one of the following:

a leucine at position 400 of SEQ ID NO 2;

10 an isoleucine at position 405 of SEQ ID NO 2;

an arginine at position 488 of SEQ ID NO 2;

a phenylalanine at position 643 of SEQ ID NO 2;

or a fragment thereof comprising at least 10 amino acids provided that the fragment comprises at least one allelic variant.

15 Fragments of polypeptide are at least 10 amino acids, more preferably at least 15 amino acids, more preferably at least 20 amino acids.

According to another aspect of the present invention there is provided an antibody specific for an allelic variant of human OATPC polypeptide as described herein.

Antibodies can be prepared using any suitable method. For example, purified polypeptide may be utilized to prepare specific antibodies. The term "antibodies" is meant to include polyclonal antibodies, monoclonal antibodies, and the various types of antibody constructs such as for example $F(ab')_2$, Fab and single chain Fv. Antibodies are defined to be specifically binding if they bind the allelic variant of OATPC with a K_a of greater than or equal to about $10^7 M^{-1}$. Affinity of binding can be determined using conventional techniques, for example those described by Scatchard et al., *Ann. N.Y. Acad. Sci.*, 51:660 (1949).

25 Polyclonal antibodies can be readily generated from a variety of sources, for example, horses, cows, goats, sheep, dogs, chickens, rabbits, mice or rats, using procedures that are well-known in the art. In general, antigen is administered to the host animal typically through parenteral injection. The immunogenicity of antigen may be enhanced through the use of an adjuvant, for example, Freund's complete or incomplete adjuvant. Following booster immunizations, small samples of serum are collected and tested for reactivity to antigen. Examples of various assays useful for such determination include those described in:

Antibodies: A Laboratory Manual, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988; as well as procedures such as countercurrent immuno-electrophoresis (CIEP), radioimmunoassay, radioimmunoprecipitation, enzyme-linked immuno-sorbent assays (ELISA), dot blot assays, and sandwich assays, see U.S. Patent Nos. 4,376,110 and 4,486,530.

- 5 Monoclonal antibodies may be readily prepared using well-known procedures, see for example, the procedures described in U.S. Patent Nos. RE 32,011, 4,902,614, 4,543,439 and 4,411,993; *Monoclonal Antibodies, Hybridomas: A New Dimension in Biological Analyses*, Plenum Press, Kennett, McKearn, and Bechtol (eds.), (1980).

- The monoclonal antibodies of the invention can be produced using alternative
 10 techniques, such as those described by Altling-Mees et al., "Monoclonal Antibody Expression Libraries: A Rapid Alternative to Hybridomas", *Strategies in Molecular Biology* 3: 1-9 (1990) which is incorporated herein by reference. Similarly, binding partners can be constructed using recombinant DNA techniques to incorporate the variable regions of a gene that encodes a specific binding antibody. Such a technique is described in Larrick et al., *Biotechnology*, 7:
 15 394 (1989).

Once isolated and purified, the antibodies may be used to detect the presence of antigen in a sample using established assay protocols, see for example "A Practical Guide to ELISA" by D. M. Kemeny, Pergamon Press, Oxford, England.

- According to another aspect of the invention there is provided a diagnostic kit
 20 comprising an antibody of the invention.

The invention will now be illustrated but not limited by reference to the following Examples. All temperatures are in degrees Celsius.

In the Examples below, unless otherwise stated, the following methodology and materials have been applied.

- 25 AMPLITAQ™, available from Perkin-Elmer Cetus, is used as the source of thermostable DNA polymerase.

General molecular biology procedures can be followed from any of the methods described in "Molecular Cloning - A Laboratory Manual" Second Edition, Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory, 1989).

- 30 Electropherograms were obtained in a standard manner: data was collected by ABI377 data collection software and the wave form generated by ABI Prism sequencing analysis (2.1.2).

Example 1**Identification of Polymorphisms****1. Methods**DNA Preparation

- 5 DNA was prepared from frozen blood samples collected in EDTA following protocol I (Molecular Cloning: A Laboratory Manual, p392, Sambrook, Fritsch and Maniatis, 2nd Edition, Cold Spring Harbor Press, 1989) with the following modifications. The thawed blood was diluted in an equal volume of standard saline citrate instead of phosphate buffered saline to remove lysed red blood cells. Samples were extracted with phenol, then
- 10 phenol/chloroform and then chloroform rather than with three phenol extractions. The DNA was dissolved in deionised water.

Template Preparation

- Templates were prepared by PCR using the oligonucleotide primers and annealing temperatures set out below. The extension temperature was 72° and denaturation temperature
- 15 94°. Generally 50 ng of genomic DNA was used in each reaction and subjected to 35 cycles of PCR. Where described below, the primary fragment was diluted 1/100 and two microlitres were used as template for amplification of secondary fragments. PCR was performed in two stages (primary fragment then secondary fragment) to ensure specific amplification of the desired target sequence.

20 Polymorphisms in OATPC: cDNA screening of 15 Liver samples

Region	SNP	Position	Amino Acid Change	Allele frequencies
Exon 4	G/A	510	None	G=96.7% A=3.3%
Exon 5	C/T	670	None	C=50% T=50%
Exon 5	C/T	696	None	C=60% T=40%
Exon 9	C/G	1299	Phe400Leu	C=96.7% G=3.3%
Exon 9	G/A	1312	Val405Ile	G=96.7% A=3.3%
Exon 9	G/A	1347	None	G=96.7% A=3.3%
Exon 10	G/C	1561	Gly488Arg	G=96.7% C=3.3%
Exon 14	A/C	2028	Leu643Phe	A=90%

				C=10%
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OATP2 above refers to the clone sequenced by Hsiang et al (ref 1). Some comment on the numbering of exons in OATPC is required. This gene contains an exon (38 bp) upstream (5' UTR region) of the exon containing the ATG start site for translation. Therefore the exon numbering could vary depending whether this exon is counted as the first exon or not. In the literature, Konig (2000) JBC 275: 23161-68, have defined exon 1 as that containing the ATG start site and therefore we have adopted the same numbering in this application (but note that the priority document relating to the present application did vice versa; for example, exon 5 in this application is equivalent to exon 6 in the priority document).

PCR PRODUCTS

Fragment	Forward Oligo	Reverse Oligo
443-999	443-466	979-999
874-1360	874-896	1337-1360
1255-1684	1255-1278	1663-1684
1559-2095	1559-1581	2073-2095

20 RFLP analysis

Polymorphism	Position	RFLP Enzyme/ PCR size	RFLP fragment size
G/A	510		
C/T	670	BmR I/ 595bp	C=349bp, 246bp T=595bp
C/T	696		
C/G	1299	Apo I/ 595bp	C=30bp, 60bp, 380bp G=90bp, 380bp
G/A	1312	Bst 4CI/ 595bp	G=77bp, 393bp A=470bp
G/A	1347		
G/C	1561	HpyCH4IV/ 595bp	G=470bp C=144bp, 326bp
A/C	2028	Ase I/ 577bp	A=89bp, 488bp C=577bp

Example 2**Further OATPC Polymorphisms**

SNPs in OATPC 3'UTR (positions according to SEQ ID NO 1)							
Exon	Nucleotide	SNP				Frequency	Frequency
						Caucasian	Japanese
3' UTR	2327	Ins T				not screened	0.1
3' UTR	2342	T>C				not screened	0.4
SNPs in OATPC promoter (positions according to SEQ ID NO 3)							
	Nucleotide	SNP				Frequency	Frequency
						Caucasian	Japanese
	321	T>G				0.03	not screened
	1332	A>C				0.08	not screened
SNPs in OATPC introns							
Intron	Nucleotide	SNP	Nucleotide	Sequence			
	position in		position	ID No			
	relation to exon		in sequence				
1	IVS1+21	T>A	41	4			
2	IVS2+89	T>G	109	5			
2	IVS2+224	A>G	244	5			
3	IVS3+97	C>A	117	6			
3	IVS3+263	G>A	283	6			
4	IVS4+189	G>A	209	7			
4	IVS4+191	G>A	211	7			
4	IVS5-118	delCTTGTA	63	8			
6	IVS6+33	C>T	53	9			
9	IVS10-107	ins TTC	75	10			
11	IVS11+142	ins T	162	11			
12	IVS13-97	G>C	84	12			

5 OATPC intronic SNPs**Key**

20bp of exon sequence shown in uppercase

Intron sequence in lowercase (200 to 300bp only)

10 SNP shown in uppercase (one allele only)

Sequence ID No 4

IVS1+21 T>A

SNP at position 41 in this sequence

15 GATACTGCAA TGGATTGAAG gtagaataag ttttatgttt Ttgagctaaa ataagtaaat 60

```

aggggaacttt aatgtataga aaagcaagtt gttaaaaaga acattatggt tcaaattata 120
attttcaatt gaagcatata ttgaaatatt aacataatga ttcataacct gattttaaacc 180
agtcttttaa tctgattaag 200

```

5

Sequence ID No 5

IVS2+89 T>G SNP at position 109 in this sequence

IVS2+224 A>G SNP at position 244 in this sequence

```

10 TGACGGAAGC TTTGAAATTG gtaacattta ttttctattt taataaccaa accctgcaag 60
   ttaaaaaata tatatgcctt acaccactgg ttatcaactg gggtaaatTt atctctcaca 120
   ggcaaatgtg caataactaa aaacatttgt ggtgtgcata actgcacagg ggttgggggc 180
   aatggaagtg ctactggtat ctaaaggtag aggtcagggg tactgctaaa tattctataa 240
   tgcAcaaaaga atgatgtaac tgaaaatggt gatagtggagg atgttcagaa accctgattc 300

```

15

Sequence ID No 6

IVS3+97 C>A SNP at position 117 in this sequence

IVS3+263 G>A SNP at position 283 in this sequence

```

20 CCACATTTCT TCATGGGATA gtaagtgtta aaaaaaaaaa aaacctctgt gccactatca 60
   tgaccttgta aattaggagt agaattttat tattatccct ttataataggc agttacCttt 120
   tgagaagata cccactaagt gtgtacacaa atgaaatagt gtctatttgt ctacataatc 180
   attttattta tctgtagctt catatacttt gaaataacaa aaagactaaa ctgtgatata 240
   tcaaatgaaa taatataggct ttttatgaat ttttagtata acGtatatac tgtacgtctt 300

```

Sequence ID No 7

IVS4+189 G>A SNP at position 209 in this sequence

IVS4+191 G>A SNP at position 211 in this sequence

```

30 ACCTGACATA GTGGGAAAAG gtaagaatta atattgcacg ta aaaagtct tctaaaaatgt 60
   atacatttaa tcatctctct aaaaatttgt gtgatattca tttagcaaat ttaattaaaga 120
   atgaatagga aaaaacatttg actcttcacag acataattat agtggttaata tacacagctc 180
   gccatttaac aacacagggt taaactacGc Gttttcactt ctatgcaaat ttgtgccatc 240
   tgaactggat gataaacctg ccggtgaaga tatctgacat tttctatatt tggattgaac 300

```

Sequence ID No 8

IVS5-118 delCTTGTA deletion of 6bp from position 63 to 68 incl.

```

40 tagcagcata agaattgact aatacaccat attgtcaaaag tttgcaaaag gaatataaaat 60
   taCTTTGACT tgtaaatttaa aaaaaataaa gtagaataat taagagttaa caagtagtta 120
   aatttggtaa agaaatgcta aaattaatgt ttaaaatgaa acactctctt atctacatag 180
   GTTGTTTAA GGAATCTGGG 200

```

45 Sequence ID No 9 IVS6+33 C>T SNP at position 53 in this sequence

```

TATTGGATAT GTAGACTTAA gtaagtacaa ccagaacaa gtaccatgat aaCgtctttc 60
taagcacaca tgcgaaaac attttttcaa ataactgaat tcactctttc aatagtcctt 120
tgcttaatat aattagaaga ttacaagtag gaaataaatg tattactaat cagaataaat 180
ataaaatcca gctctctatt 200

```

50

Sequence ID No 10

IVS10-107 Ins TTC SNP at position 75 in this sequence

```

55 ttaaaaaaaa ctttgccatt tctgtcatcat caaagcaaat ttcttcatat aagaaaaaat 60
   tctttatcta cttt(TTC)ttttc ctctttctct gctttcactt tactctctcc ttctctccc 120
   ctctttgttc ttttctctct ctctctctct ttttgatata tgtctatcat atatttcag 180
   AATAATCCA GTGACATCTC 200

```

Sequence ID No 11

IVS11+142 InsT InsT at position 162 in this sequence

```

60 CATGTCATGC TGATTGTTAA gtaagtatga cttttaaaaa cattttcata tgcattgagac 60
   tataaacaca cctaattgata tgcattatatt tacataatat actgggaatt caaattcata 120
   tttcatcaaa ttttaatttt ctgagaattc attttattaa aa(T)ttactatg aactctcaag 180
   gctgtaatta ataattttgc 200

```

Sequence ID No 12

IVS13-97 G>C

SNP at position 84 in this sequence

5 tgatttgggt ctttgagatt tctaataate tttattattg ggtagatgca gaacaaaaa 60
 ataaacgaat cctccaaatt tttGaaacttt tatTTaataca aaatatataa atgtggaata 120
 tcatgcagtt acatttataaa tatgttccct aaactgacat cttctctctt cctattacag 180
 GAGGAATTCT AGCTCCAATA 200

OATPC promoter region

Total length of the sequence = 1538bp

1500bp of OATPC sequence directly upstream from the cDNA sequence

Sequence in uppercase represents 38bp overlap with the cDNA sequence (SEQ ID NO 1)

15 where this 38bp is 5'UTR sequence.

Nucleotide positions in the promoter have been determined where the -1 position is the base (lowercase) directly upstream of the end of the cDNA sequence.

20 atgctctttg acctctgaaa atattggaga attttacaac tggcaccttt agctcaggat 60
 tataaaaggt gttagttagt ttgtactgtt ttattcttcat tglataataat atatatatta 120
 gtctccaaac atgttgatgt gttttcaatg aaatggatgt ctgaggagaa aacctattagc 180
 ctgagaaaaa ccaactgta ttccattgtt gaataaaagg aagtcataaa aaatgatgga 240
 aaatgttctg cattcctgtt atgatatcaa aatctggcag tacatgaaaa tttttcaaaag 300
 25 tgcttattta acaggcataa tctttggtct cctgagccag aatctgctgg gtatgggact 360
 ggattgctat ttgacaact cggcagtaga ttcttaacta gcagagtatt tggaaagcctt 420
 actctaatat ttggcctgt ggtctacatt tctcagttct gcacagtcat tcttccctct 480
 tacactactc tttagtttgt ctcatgattc caatactctc aataaataac caagaataga 540
 actaatcaat cagataactg tggcacagac atcaaatata ttttgctgca accatatcaa 600
 caaatgtccc atgaatgata aggggtaacc atattctcat atatgcattc tcacattacc 660
 30 acatatatat atgtgcatat gtgtatacag gtaaaagtgt gtatatatgt atacatgtat 720
 gtttgtgtgt alatacatcac atatatcttc acacttttct gaaatatata tattttatgtg 780
 agagaagggt ctgtacttta ttccagaaga gagcttaagt tccaaggtat aattgagaggt 840
 ctaaaatgtt tgagttattg aatcaalttaa acttcatctc tactcaagaa aacttttaac 900
 tgagttaaag tcttctcttc tcccaagtc aagccaataa aaggaaaactg tgatattaat 960
 35 aattcttctc tgttttgatg taaagaatct gcagttctta ttttcatcat 1020
 tcagaaaaat ggtcttgacg ttaattggga ctctcttatt ccagggtgga tctccagttct 1080
 ccatacatcac cactgttagaa ccatacttat gtaccaagca aagagggtat attttaattct 1140
 ttaaatgccca atgtaacctg taggcataat ttttatttgt cttaaatatt tctctatttg 1200
 gaagttttaa atacctggaa taatttattg tactcatatt tttaaagaaa aaatcttat 1260
 40 gccaccaact taattgaata aacaagtaaa agccattccc aaaagtaagg ttactctgtt 1320
 aagattaaaca aaaaataatg tgagaattct gagaataata atctttaaat attggcaact 1380
 ggaagtgaact cttaaaaacta actaggtttt atatgtttga ctagagcaat gacataataa 1440
 ggtggttaat catcactgga cttgttttca aaaagccaac tacttaaga ggaataaagg 1500
 GTGGACTTGT TGCAATTGCT GTAGATTCT AAATCCAG 1538